

Effects of Vitamin E on Receptor Activator of Nuclear Factor Kappa B Ligand (RANKL) and Osteoprotegerin (OPG) in Rats Treated With Nicotine

M Norazlina, PhD*, J Maizatul-Neza, BSc**, A Azarina, BSc**, A S Nazrun, PhD*, M Norliza PhD*, S Ima-Nirwana, PhD*

*Department of Pharmacology, Faculty of Medicine and **Department of Biomedical Sciences, Faculty of Allied Health Sciences, Universiti Kebangsaan Malaysia, Jalan Raja Muda Abdul Aziz, 50300 Kuala Lumpur, Malaysia

SUMMARY

Vitamin E is found to reverse the effects of nicotine on bone and this study aimed to determine its mechanism. Male Sprague Dawley rats were divided into four groups and treated for 3 months: Group 1 was the control group (RC). Groups 2 (N), 3 (N+TT) and 4 (N+ATF) received nicotine 7 mg/kg throughout the treatment period. In addition, groups 3 and 4 received tocotrienol 60 mg/kg and α -tocopherol 60 mg/kg respectively during months 2 and 3. Parameters measured were serum osteoprotegerin (OPG), serum receptor activator of nuclear factor kappa B ligand (RANKL), femoral and lumbar bone calcium content and body weight. Nicotine did not affect OPG or RANKL levels but reduced bone calcium content suggesting the calcium loss is not due to increase osteoclastogenesis. OPG was increased in N+ATF while RANKL was slightly increased in N+TT. Both vitamin E supplements restored bone calcium loss induced by nicotine. Nicotine impaired weight gain in all treatment groups starting week 4 however, N+TT group was comparable to RC from week 6 onwards. Bone protective effects of ATF, but not TT, may be partly due to inhibition of osteoclastogenesis.

KEY WORDS:

Nicotine, Vitamin E, Osteoprotegerin, RANKL ligand, Rats

INTRODUCTION

Bone is a specialized connective tissue which undergoes constant remodeling; composed of bone resorption and bone formation processes. It is possible to measure the remodeling processes by means of various biochemical markers.

Osteoprotegerin (OPG) is a protein which protects bone from being degraded¹. It is also known as osteoclast inhibitory factor (OCIF)². It is secreted by osteoblasts³ and acts as a false receptor for receptor activator of nuclear factor kappa B ligand (RANKL) which leads to prevention of production and activation of osteoclasts⁴. Osteoprotegerin has also been reported to stimulate apoptosis of osteoclasts⁵.

RANKL, on the other hand, is a molecule produced by osteoblasts which has an important role in the differentiation of osteoclasts⁵. Binding of RANKL to its receptor, RANK, stimulates osteoclastogenesis in vitro which requires the presence of macrophage-colony stimulating factor (M-CSF)⁶. Alterations in the ratio of RANKL to OPG may lead to several skeletal-related diseases⁷.

Nicotine has been shown to increase the production of inflammatory mediators⁸ and nicotine has been associated with pathogenesis of diseases by promoting proinflammatory mediators⁹. On bone, nicotine induced bone loss and reduced bone mechanical strength in rats¹⁰. In humans, smoking has been implicated in osteoporosis¹¹ and is considered one of the risk factors for osteoporosis¹². Our previous studies showed that nicotine increased bone resorbing cytokines, interleukin 1 and 6¹³ and impaired bone histomorphometrically¹⁴. Interestingly, vitamin E, a naturally-occurring antioxidant, was able to prevent the increment of bone resorbing cytokines¹⁵ and reverse the damage on bone histomorphometry¹⁶ in nicotine-induced rats.

This study was carried out to further determine the mechanisms of how nicotine damages the bone by measuring OPG and RANKL levels as well as bone calcium content. In addition, the effects of vitamin E (tocotrienol mixture and α -tocopherol) on the above parameters were also determined.

MATERIALS AND METHODS

Animals and treatment

Three-month-old male Sprague-Dawley rats obtained from the Laboratory Animal Resource Unit, Universiti Kebangsaan Malaysia, were randomly divided into four groups of eight rats each: (1) control group (RC), (2) nicotine-treated group at the dose of 7 mg/kg for 3 months (N), (3) group treated with nicotine (7 mg/kg) for 3 months and supplemented with tocotrienol mixture (60 mg/kg) during months two and three (N+TT) and (4) group treated with nicotine (7 mg/kg) for three months supplemented with α -tocopherol (60 mg/kg) during months two and three (N+ATF). Another group of eight rats were sacrificed untreated and acted as baseline control (BC).

The rats were kept four per cage under 12-hour natural light/dark cycles, given tap water *ad libitum* and weighed every week. All rats received normal rat chow obtained from Gold Coin (Port Klang, Selangor, Malaysia). The nicotine used was in the form of hydrogen tartrate salt which was purchased from Sigma Chemical Co. (St Louis, MO, USA). Alpha-tocopherol acetate was purchased from Sigma Chemical Co. (St Louis, MO, USA) while the Malaysian Palm Oil Board (Bangi, Selangor, Malaysia) supplied the tocotrienol mixture. Blood samples were obtained before the treatment commenced and at the end of the treatment period.

This article was accepted: 15 January 2010

Corresponding Author: Norazlina Mohamed, Department of Pharmacology, Faculty of Medicine, Universiti Kebangsaan Malaysia, Jalan Raja Muda Abdul Aziz, 50300 Kuala Lumpur, Malaysia Email: azlina@medic.ukm.my

Nicotine (7 mg/kg) was prepared by mixing 0.07 g of nicotine in 10 ml normal saline. The vitamin E solution was prepared by mixing 3 g of the respective vitamin Es in 50 ml olive oil (Bertolli, Secaucus, NJ, USA). A total of 0.1 ml/100 g rat weight of the nicotine and vitamin E preparations were given respectively intraperitoneally and orally via oral gavage, six days a week. The control group received the vehicles i.e. normal saline, intraperitoneally, and olive oil, via oral gavage. Upon sacrifice of the rats, left femur and 4th lumbar vertebra bones were harvested for bone calcium content analysis.

Bone biochemical markers

Serum OPG levels were measured using ELISA kit Cat No BI-20602 while serum RANKL levels were determined using ELISA kit Cat No BI-20522. Both kits were purchased from Biomedica Medizinprodukte GmbH Co, Vienna, Austria.

Bone calcium content

For the measurement of bone calcium content, briefly, the femur and 4th lumbar bones were subjected to two phases of drying i.e. 100°C for 24 hours and 800°C for 12 hours. The bones were then dissolved in nitric acid and the solutions were diluted with lanthanum chloride before analysis was carried out using flame atomic absorption spectrophotometry (Analyst 100, Perkin ElmerTM Instruments, Wellesley, Massachusetts, USA) at 422.7 nm.

Analyses of data

All the data were found to be normally distributed by the Kolmogorov-Smirnov test. The data were then analysed using the one-way analysis of variance (ANOVA) and Tukey's honestly significant difference test was selected as the post-hoc test. To compare data before and after treatment, the paired student's t-test was used. All the analysis was carried out using the Statistical Package for Social Sciences version 12.0 (SPSS Inc, Chicago, IL, USA) software.

This study was approved by the Universiti Kebangsaan Malaysia Animal Ethics Committee (UKMAEC) with the approval number FAR/2003/IMA/12JUNE/095.

RESULTS

Serum OPG levels did not show any significant difference in nicotine-treated group as compared to the control group (Fig 1). Even though nicotine group showed a trend of lowering the OPG levels, significance level was not reached. Supplementing the rats with vitamin E (tocotrienol mixture and α -tocopherol) increased OPG levels but the values did not differ as compared to RC and N groups. However, OPG level in N+ATF group after the treatment was significantly different than before treatment.

Similar to OPG, RANKL levels were not different when compared between the RC and N groups (Fig 2). However, a trend of increasing RANKL levels could be observed in N group. No changes were observed in the vitamin E supplemented groups as compared to RC and N groups. However, N+TT group had a higher level of RANKL after treatment than before treatment.

Femur bone calcium content was reduced in nicotine treated group as compared to RC group (Fig 3). Supplementation of tocotrienol and tocopherol were able to reverse the effects and bone calcium content was increased back to normal

levels and was higher compared to BC group. In addition, femur bone calcium content of the N+ATF was even higher than the RC group. No significant changes were observed in the 4th lumbar bone calcium content amongst the four treatment groups except that all four groups had a higher bone calcium content than the BC group (Fig 3).

The body weight of the rats showed an increasing trend from week 0 to week 12 for all groups (Fig 4). Starting from week 3, the RC group had a higher body weight than N+TT and N+ATF. At week 4, body weight of RC group was significantly higher than the other treatment groups. The trend persisted until the end of the treatment period except for N+TT which had comparable body weight to RC starting from week 6.

DISCUSSION

The dose of nicotine used in this study (7 mg/kg) was based on an earlier study i.e. the effective dose in causing an increase in bone resorbing cytokines¹⁷ and reducing bone calcium content¹⁸.

In this study, administration of nicotine for three months did not cause any changes in the OPG levels. We also observed that nicotine did not cause an increase in RANKL levels. OPG and RANKL are indicative of osteoclastogenesis inhibition and stimulation respectively. To the best of our knowledge there is paucity of reports on nicotine and OPG or RANKL. In an in vitro study, administration of nicotine and lipopolysaccharide into osteoblast cell culture caused an initial increase in OPG expression but decreased at later stage of culture¹⁹.

Despite the lack of changes observed in OPG and RANKL levels, we observed bone calcium loss in the femur of the nicotine-treated group. The findings were consistent with a previous study which showed that nicotine caused a reduction in bone mineral content²⁰. However, in this study, fourth lumbar bone calcium content was not affected by nicotine treatment. Even though lumbar bones consisted of trabecular bone and was more prone to change compared to cortical bones²¹, reduction in calcium content of the lumbar bones was not observed in this study which was in contrast to our previous study¹⁸. This may be due to the fact that lumbar bones are smaller than femur bones, thereby containing less calcium. Therefore smaller changes would be more difficult to detect.

Previously, nicotine was found to stimulate osteoclast differentiation and cause resorption of calcium phosphate²². One of our earlier studies had shown that nicotine increased bone resorbing cytokines i.e. interleukin 1 and 6¹³ as well as increasing pyridinoline levels, a marker for bone resorption¹⁴. The above findings suggested that nicotine induces bone loss via other mechanisms and not through stimulation of osteoclastogenesis.

Supplementation of α -tocopherol to the nicotine-treated rats caused an increase in OPG levels. Since nicotine did not affect OPG levels, the changes observed upon α -tocopherol supplementation may be due to the supplementation itself. Although there has been no report on the effects of vitamin E on OPG levels, other antioxidants have been found to affect OPG levels and its mRNA expression^{23,24}. Increase in OPG levels implied reduction in bone resorption activity which

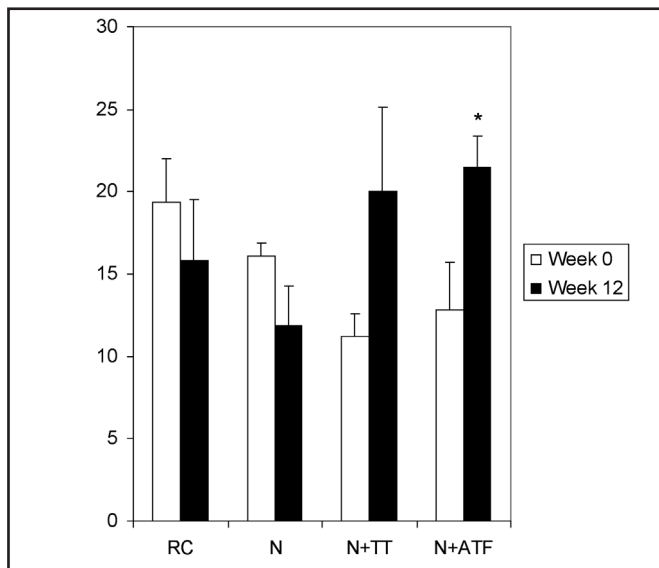


Fig. 1: Effects of nicotine administration and vitamin E supplementation on serum OPG levels.

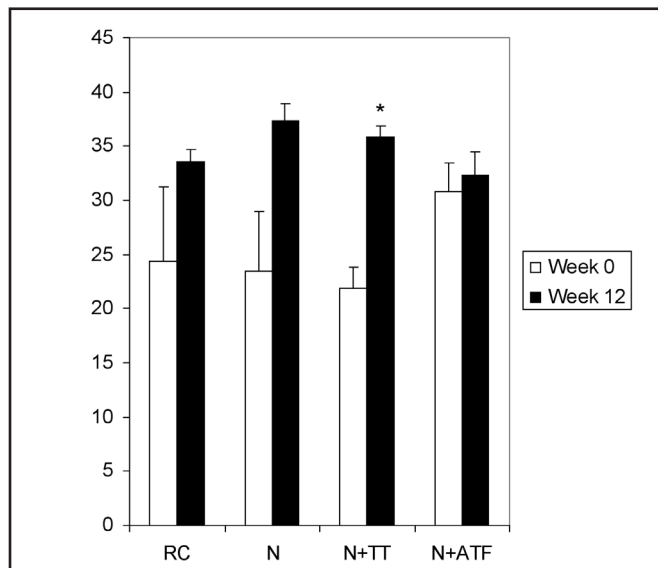


Fig. 2: Effects of nicotine administration and vitamin E supplementation on serum RANKL levels.

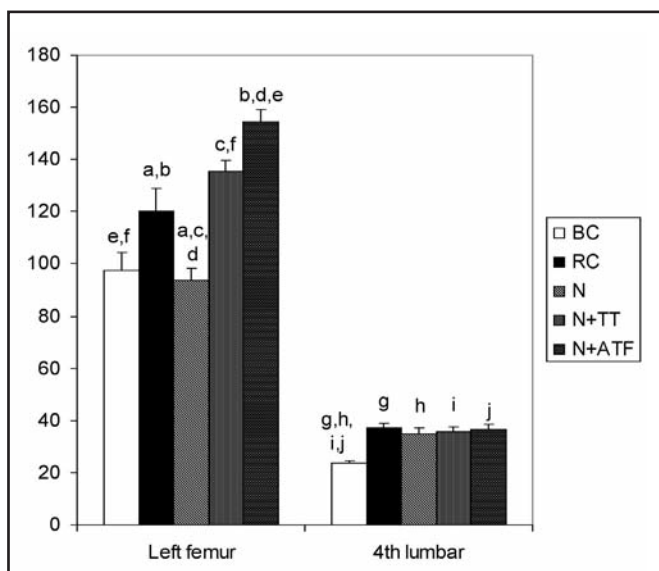


Fig. 3: Effects of nicotine administration and vitamin E supplementation on femoral and lumbar bone calcium content.

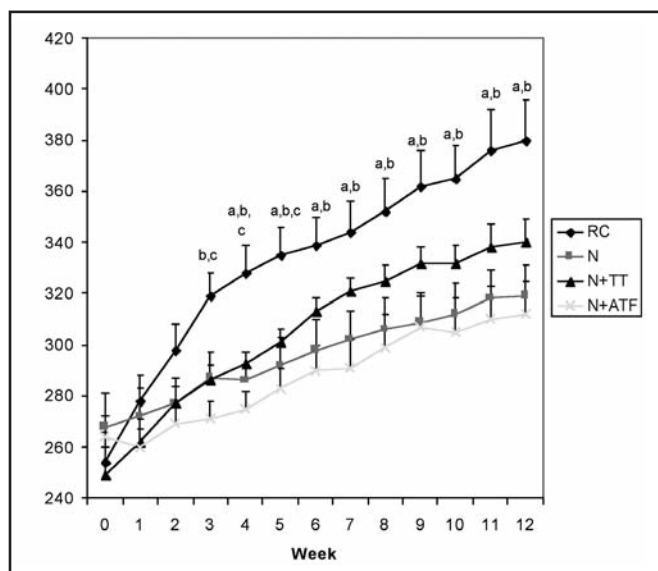


Fig. 4: Effects of nicotine administration and vitamin E supplementation on body weight.

Guidelines to Figures 1-3

- BC - baseline control
- RC - rat chow
- N - nicotine 7 mg/kg body weight
- N+TT - nicotine 7mg/kgbody weight (1 month) followed by tocotrienol 60 mg/kg body weight (2 months)
- N+ATF - nicotine 7mg/kgbody weight (1 month) followed by α -tocopherol 60 mg/kg body weight (2 months)
- * indicates significant difference to week 0 ($p < 0.05$)
- Groups which share the same alphabet indicates significant difference ($p < 0.05$)

Guidelines to Figure 4

- BC - baseline control
- RC - rat chow
- N - nicotine 7 mg/kg body weight
- N+TT - nicotine 7mg/kgbody weight (1 month) followed by tocotrienol 60 mg/kg body weight (2 months)
- N+ATF - nicotine 7mg/kgbody weight (1 month) followed by α -tocopherol 60 mg/kg body weight (2 months)
- a, b and c indicate significant difference between RC and N, RC and N+ATF and RC and N+TT respectively ($p < 0.05$)

would then lead to prevention of bone loss, hence the reversal of bone calcium loss seen in this group. This effect was not observed in the N+TT group.

The N+TT group did not show any change in OPG levels. Instead the RANKL levels were higher than before treatment. Again, since nicotine did not affect RANKL levels, we suggest that the tocotrienol mixture itself caused the increase in RANKL levels. To date, no reports have been published on the effects of vitamin E on RANKL. However, one study using α -lipoic acid, an antioxidant, reported an up-regulation of RANKL production by α -lipoic acid²⁵. In that study, the researchers observed that even though RANKL was increased, osteoclastogenesis and bone resorption were reduced which led them to suggest that α -lipoic acid inhibited osteoclastogenesis by inhibiting RANKL signals and not by modulating RANKL production.

Similar arguments may be applied in the present study. Even though we found that RANKL was increased in N+TT group, the group was able to prevent bone calcium loss due to nicotine. We have also shown that tocotrienol, especially the gamma isomer, prevent adverse effects of nicotine in terms of biochemical markers (unpublished data) and histomorphometric parameters¹⁶. In other studies, tocotrienol was able to prevent bone loss in orchidectomized rats²⁶ and maintain bone mineral density in ovariectomized rats²⁷. This further suggested that the effects of tocotrienol in reversing nicotine-induced bone loss was not via the modification of OPG or RANKL activities. Other mechanisms might be responsible for the effects of tocotrienol.

In terms of body weight, this study is consistent with previous study which found reduction in body weight of rats after administration of nicotine²⁸. It was suggested that nicotine affected fat stores in the body. In another study, Schwid *et al.* reported that nicotine-treated rats experienced hypophagia thus leading to weight loss²⁹. The reduction in food intake in rats exposed to nicotine is due to involvement of perifornical hypothalamus through the action of catecholaminergic neurons³⁰. Vitamin E, on the other hand, may play a role in maintaining body mass. Our previous study showed that rats fed with vitamin E deficient diet had a declining body weight as compared to the control rats³¹. In the present study, we observed that nicotine-treated rats supplemented with tocotrienol mixture were able to reverse the effects of nicotine on weight gain.

In conclusion, nicotine induced bone resorption without affecting OPG or RANKL levels. Nicotine also caused growth retardation. Both tocotrienol and α -tocopherol, via separate mechanisms, were able to restore bone calcium content which was lost due to nicotine treatment. However, tocotrienol supplementation, but not α -tocopherol, improved growth in nicotine-treated rats.

REFERENCES

- William JB, David LL. Osteoprotegerin In: Ernesto C (ed). Skeletal growth factor. Philadelphia: Lippincott Williams & Wilkins, 2000; 365-74.
- Wong BR, Josien R, Choi Y. TRANCE is a TNF family member that regulates dendritic cell and osteoclast function. *J Leukoc Biol* 1999; 65: 715-24.
- Boyce BF, Xing L. The RANKL/RANK/OPG pathway. *Curr Osteoporos Rep* 2007; 5: 98-104.
- Hofbauer LC, Khosla S, Dunstan CR, Lacey DL, Boyle WJ, Riggs BL. The roles of osteoprotegerin and osteoprotegerin ligand in the paracrine regulation of bone resorption. *J Bone Miner Res* 2000; 15: 2-12.
- Shiotani A, Takami M, Itoh K, Shibasaki Y, Sasaki T. Regulation of osteoclast differentiation and function by receptor activator of NF κ B ligand and osteoprotegerin. *Anat Rec* 2002; 268: 137-46.
- Quinn JM, Elliott J, Gillespie MT, Martin TJ. A combination of osteoclast differentiation factor and macrophage colony stimulating factor is sufficient for both human and mouse osteoclast formation in vitro. *Endocrinol* 1998;139: 4424-27.
- Vega D, Maalouf NM, Sakhaee K. Clinical Review #: the role of receptor activator of nuclear factor-kappaB (RANK)/RANK ligand/osteoprotegerin: clinical implications. *J Clin Endocrinol Metab* 2007; 92: 4514-21.
- Jaimes EA, Tian RX, Joshi MS, Raij L. Nicotine augments glomerular injury in a rat model of acute nephritis. *Am J Nephrol* 2009; 29: 319-26.
- Cooper RG, Magwere T. Nitric oxide-mediated pathogenesis during nicotine and alcohol consumption. *Indian J Physiol Pharmacol* 2008; 52: 11-8.
- Broulik PD, Rosenkrancova J, Ruzicka P, Sedlacek R, Kurcova I. The effect of chronic nicotine administration on bone mineral content and bone strength in normal and castrated male rats. *Horm Metab Res* 2007; 39: 20-4.
- Kapoor D, Jones TH. Smoking and hormones in health and endocrine disorders. *Eur J Endocrinol* 2005; 152: 491-9.
- Chapurlat R. Epidemiology of osteoporosis. *J Soc Biol* 2008; 202: 251-5.
- Hapidin H, Othman F, Soelaiman IN, Shuid AN, Luke DA, Mohamed N. Negative effects of nicotine on bone-resorbing cytokines and bone histomorphometric parameters in male rats. *J Bone Miner Metab* 2007; 25: 93-8.
- Hermizi H, Faizah O, Ima-Nirwana S, Ahmad Nazrun S, Luke DA, Norazlina M. Nicotine impaired bone histomorphometric parameters and bone remodeling biomarkers in Sprague-Dawley male rats. *Annals of Microscopy* 2007; 7: 10-24.
- Norazlina M, Lee PL, Lukman HI, Nazrun AS, Ima-Nirwana S. Effects of vitamin E supplementation on bone metabolism in nicotine-treated rats. *Singapore Med J* 2007; 48: 195-99.
- Hermizi H, Faizah O, Ima-Nirwana S, Ahmad Nazrun S, Norazlina M. Beneficial effects of tocotrienol and tocopherol on bone histomorphometric parameters in Sprague-Dawley male rats after nicotine cessation. *Calcif Tissue Int* 2009; 84: 65-74.
- Norazlina M, Nik-Farideh YMK, Arizi A, Faisal A, Ima-Nirwana S. Effects of nicotine on bone resorbing cytokines in male rats. *Int Med J* 2004; 3: <http://www.e-imj.com/Vol3-No2/Vol3-No2-B10.htm>
- Ima-Nirwana S, Cheng CT, Norazlina M. Effects of nicotine on bone mineral density and calcium homeostasis in male Sprague-Dawley rats. *Current Topics in Pharmacology* 2005; 9: 125-29.
- Tanaka H, Natsuko T, Maiko S *et al.* Nicotine and lipopolysaccharide stimulate the formation of osteoclast-like cells by increasing macrophage colony-stimulating factor and prostaglandin E2 production by osteoblasts. *Life Sci* 2006; 78: 1733-40.
- Broulik PD, Rosenkrancova J, Ruzicka P, Sedlacek R, Kurcova I. The effect of chronic nicotine administration on bone mineral content and bone strength in normal and castrated male rats. *Horm Metab Res* 2007; 39: 20-4.
- Buckwalter JA, Glimcher MJ, Cooper RR, Recker R. Bone biology, Part II: Formation, form, modeling, remodeling and regulation of cell function (Instructional course lectures). *Bone Joint Surg Am* 1995; 77-A: 1276-89.
- Henemyre CL, Scales DK, Hokett SD *et al.* Nicotine stimulates osteoclast resorption in porcine marrow model. *J Periodontol* 2003; 74: 1440-46.
- Son E, Do H, Joo HM, Pyo S. Induction of alkaline phosphatase activity by L-ascorbic acid in human osteoblastic cells: a potential role for CK2 and Ikaros. *Nutrition* 2007; 23: 745-53.
- Franklin M, Bu SY, Lerner MR *et al.* Dried plum prevents bone loss in a male osteoporosis model via IGF-I and the RANK pathway. *Bone* 2006; 39: 1331-42.
- Koh JM, Lee YS, Byun CH *et al.* Alpha-lipoic acid suppresses osteoclastogenesis despite increasing the receptor activator of nuclear factor kappaB ligand/osteoprotegerin ratio in human bone marrow stromal cells. *J Endocrinol* 2005; 185: 401-13.
- Ima-Nirwana S, Kiftiah A, Zainal AG, Norazlina M, Gapor MT, Khalid BAK. Palm vitamin E prevents osteoporosis in orchidectomized growing male rats. *Natural Product Sciences* 2000; 6: 155-60.
- Norazlina M, Ima-Nirwana S, Gapor MT, Khalid BAK. Palm vitamin E is comparable to α -tocopherol in maintaining bone mineral density in ovariectomised female rats. *Exp Clin Endocrinol Diab* 2000; 108: 305-10.
- Winders SE, Grunberg NE. Effects of nicotine on body weight, food consumption and body composition in male rats. *Life Sci* 1990; 46: 1523-30.
- Schwid SR, Hirvonen MD, Keesey RE. Nicotine effects on body weight: a regulatory perspective. *Am J Clin Nutr* 1992; 55: 878-84.
- Kramer PR, Guan G, Wellman PJ, Bellinger LL. Nicotine's attenuation of body weight involves the perifornical hypothalamus. *Life Sci* 2007; 81: 500-8.
- Ima-Nirwana S, Norazlina M, Abd Gapor MT, Khalid BAK. Vitamin E deficiency impairs weight gain in normal and ovariectomised growing female rats. *Medical J Islamic Academy Sci* 1998; 11: 99-105.